

2.0X Taq Master Mix, Clear (1.5mM MgCl₂) Cat #: 42-134B

Contents: 1000 Reactions

Storage: -20°C. Reagent for *in vitro* laboratory use only

General Description

Apex Taq DNA Polymerase Master Mix, Clear from Genesee Scientific is a ready-to-use 2.0X reaction mix. Simply add primers, template, and water to successfully carry out primer extensions and other molecular biology applications.

Apex Taq DNA Polymerase, an optimized NH₄⁺ buffer system, dNTPs and magnesium chloride are present in the Taq DNA Polymerase Master Mix, Clear. Each reaction requires 25 μ l of the 2.0X reaction mix. Simply add primers, template and water to a total reaction volume of 50 μ l.

Taq DNA Polymerase Master Mix, Clear offers several advantages. Specificity is increased. Set up time is significantly reduced. The chance of contaminating component stocks is eliminated. Reduction of reagent handling steps leads to better reproducibility. Standard tests can be set up with the confidence that results will be consistent every time.

Composition of the Taq Master Mix, Clear (2.0X)

- Tris-HCl pH 8.5, (NH₄)₂SO₄, 3 mM MgCl₂, 0.2 % Tween[®] 20
- 0.4 mM of each dNTP
- 0.08 units/µl Taq DNA polymerase

Storage and Stability

Long term storage at -20 °C. Product expiry at -20 °C is stated on the label. Option: Store at +4 °C for up to 6 months.

Quality Control

Taq DNA Polymerase is tested for contaminating activities, with no traces of endonuclease activity, nicking activity or exonuclease activity.

Protocol

This protocol serves as a guideline for primer extensions. Optimal reaction conditions such as incubation times, temperatures, and amount of template DNA may vary and must be individually determined.

Notes:

- Set up reaction mixtures in an area separate from that used for DNA preparation or product analysis.
- The table below shows the reaction set up for a final volume of 50 μL.
- Keep all components on ice.
- **Important:** Mix the solutions completely before use to avoid localized concentrations of salts.
- Vol./Reaction Final Conc. Component 2.0X Taq Master Mix, 25 μL 1X Clear Primer A Variable 0.1–1.0 μM Primer B Variable 0.1–1.0 μM Nuclease-Free Water Variable - - - -**Template DNA** Variable Variable 50 µL - - - -**TOTAL volume**
- 1. Set up each reaction as follows:

- 2. Mix gently by pipetting the solution up and down a few times.
- 3. Program the thermal cycler according to the manufacturer's instructions.

For maximum yield and specificity, temperatures and cycling times should be optimized for each new template target or primer pair.

Table 3. Three-step PCR program

Cycles	Duration of cycle	Temperature
1	2-5 minutes	95 °C
25-35	20 – 30 seconds ^a 20-40 seconds ^b 30 seconds ^c	95 °C 55 − 60 °C 72 °C
1	5 minutes ^d	72 °C

- ^{a.} Denaturation step: This step is the first regular cycling event and consists of heating the reaction to 95 °C for 20 – 30 seconds. It causes melting of the DNA template by disrupting the hydrogen bonds between complementary bases, yielding single-stranded DNA molecules.
- ^{b.} Annealing step: The reaction temperature is lowered to 50-65 °C for 20-40 seconds allowing annealing of the primers to the single-stranded DNA template. Typically, the annealing temperature is about 3-5 °C below the T_m (melting temperature) of the primers used.
- ^{c.} Extension/elongation step: Taq polymerase has its optimum activity temperature at 72 °C. At this step the DNA polymerase synthesizes a new DNA strand complementary to the DNA template strand. The extension time depends on the length of the DNA fragment to be amplified. As a rule of thumb, at its optimum temperature the DNA polymerase will polymerize a thousand bases per minute.
- ^{d.} Final elongation: This single step is occasionally performed at a temperature of 72 °C for 5 minutes after the last PCR cycle to ensure that any remaining single-stranded DNA is fully extended.

Related Products

Taq Polymerase kits (500 units)	Cat#		
With 10X Standard and Ammonium Reaction Buffer	42-800B1		
With 10X Combination Buffer	42-800B3		
Glycerol Free	42-800B4		
Hot Start DNA Polymerase (500 units)	Cat#		
With 10X Ammonium and Combination Reaction Buffer	42-106		
All polymerases are also available in kits, Mg^{2+} free buffers and 50 mM $MgCl_2$.			
Master Mixes (500 reactions)	Cat#		
2X Taq RED Master Mix, 1.5 mM MgCl ₂	42-138		
2X Taq Master Mix, Clear, 1.5 mM MgCl ₂	42-134		
2X Hot Start Master Mix Buffer I, 1.5 mM MgCl ₂	42-198		
2X Hot Start Master Mix Buffer I Blue, 1.5 mM MgCl ₂	42-144		
The shown Hot Start master mixes are ammonium based. Also available with balanced ammonium and potassium based buffers.			
Real-time PCR (400 reactions)	Cat#		
qPCR 2X Master Mix for Probe, without ROX [™]	42-116P		
qPCR 2X Master Mix for Probe, low ROX^{TM}	42-118P		
qPCR 2X Master Mix for Probe, high ROX [™]	42-120P		

42-116PG

42-118PG

42-120PG

qPCR 2X GREEN Master Mix, without ROX[™]

qPCR 2X GREEN Master Mix, low ROX[™]

qPCR 2X GREEN Master Mix, high ROX[™]

DNA Ladders	Cat#
Apex 100 bp-Low DNA Ladder, 250 applications	19-109
Apex 1 kb DNA Ladder, 333 applications	19-115
Apex 200 bp DNA Ladder, 200 applications	19-111
ECON Mini DNA Ladder 100-500 bp, 100 applications	19-130
ECON Low DNA Ladder 100-1000 bp, 100 applications	19-131
ECON PCR Ladder 100-3000 bp, 100 applications	19-132

Ultrapure dNTPs	Cat#	
dNTP set, 100 mM each:	42-410	
250 μl of each dA, dC, dG and dT	42-410	
dNTP Set, 100 mM each:	42 402	
1 ml of each dA, dC, dG and dT	42-403	
dNTP Mix 40 mM (1 x 500 μl):	42 411	
10 mM each dA, dC, dG, dT	42-411	
dNTP Mix 100 mM (2 x 1 ml):	42.405	
25 mM each dA, dC, dG, dT	42-405	
dNTP Mix 10 mM (10 x 1 ml):	12 106	
2.5 mM each dA, dC, dG, dT	42-406	
Other concentrations and Single dNTPs are available		

Other concentrations and Single dNTPs are available

Accessory reagents	Cat#
50 mM MgCl ₂ , 3 × 1.5 ml	42-303
Nuclease-Free Water, PCR Grade, 6 x 5 ml	42-710

Inspiration for Fast PCR

Fast PCR protocol: Apex Taq DNA Polymerases and Taq DNA master mixes					
90 minutes	PCR program for 3-step Standard PCR				
	Cycler step	Temp.	Duration	Cycles	
	Initial heating	95 °C	3 min.	1	
95 °C 72 °C	Denaturation	95 °C	30 sec.		
60 °C	Annealing*	60 °C	30 sec.	30	
	Extension	72 °C	30 sec.		
Standard PCR protocol	Final	72 °C	5 min.	1	
	* the annealing tempe	erature depende	s on the primer set.		
SAVE 1 HOUR	- just change PCR cycler settings!				
31 minutes	PCR program for 2-step Fast PCR				
00 %	Cycler step	Temp.	Duration	Cycles	
98 .	Initial heating	98 °C	40 sec.	1	
92 °C 72 °C	Denaturation	92 °C	2 sec.		
60 °C	Extension*	60 °C	2 sec.	30	
	Final	72 °C	5 min.	1	
2-sten Fast PCR protocol	extension * the extension temperature depends on the primer set				
2-3100 1 831 1 61 01010001	For fast PCR choose highest possible T _m .				
λ DNA Tag DNA Polymerase	Amplification	of λ DNA	-		
Taq DNA Polymerase					
STANDARD PAST					
-	Reaction mix*	•			
	Ammonium b	uf.	1x		
	dNTP mix	0	,2 mM each		
00 bp -	Primers		1,5 mivi		
) DNA		0,2 µW		
90 min 31 min			1 ng		
	Taq DNA pol.		0.5 - 10		
	* H₂O up to a to	tal volume of 25	5 µl		
Genomic DNA Tag Master Mix, Clear	Amplification	of gDNA	-		
raq master mix, oldar	2x Taq Maste	r Mix, Cle	ar		
STANDARD FAST					
-	Reaction mix*	•			
	Taq MM Clear	r	1x		
	Primers		0,2 μM		
93 bp -	gDNA		20 ng		
	* H₂O up to a to	tal volume of 25	5 µl		
90 min 31 min					